

# Promoting Fracture Healing Through Systemic or Local Administration of Allogeneic Mesenchymal Stem Cells

**Gang Li**

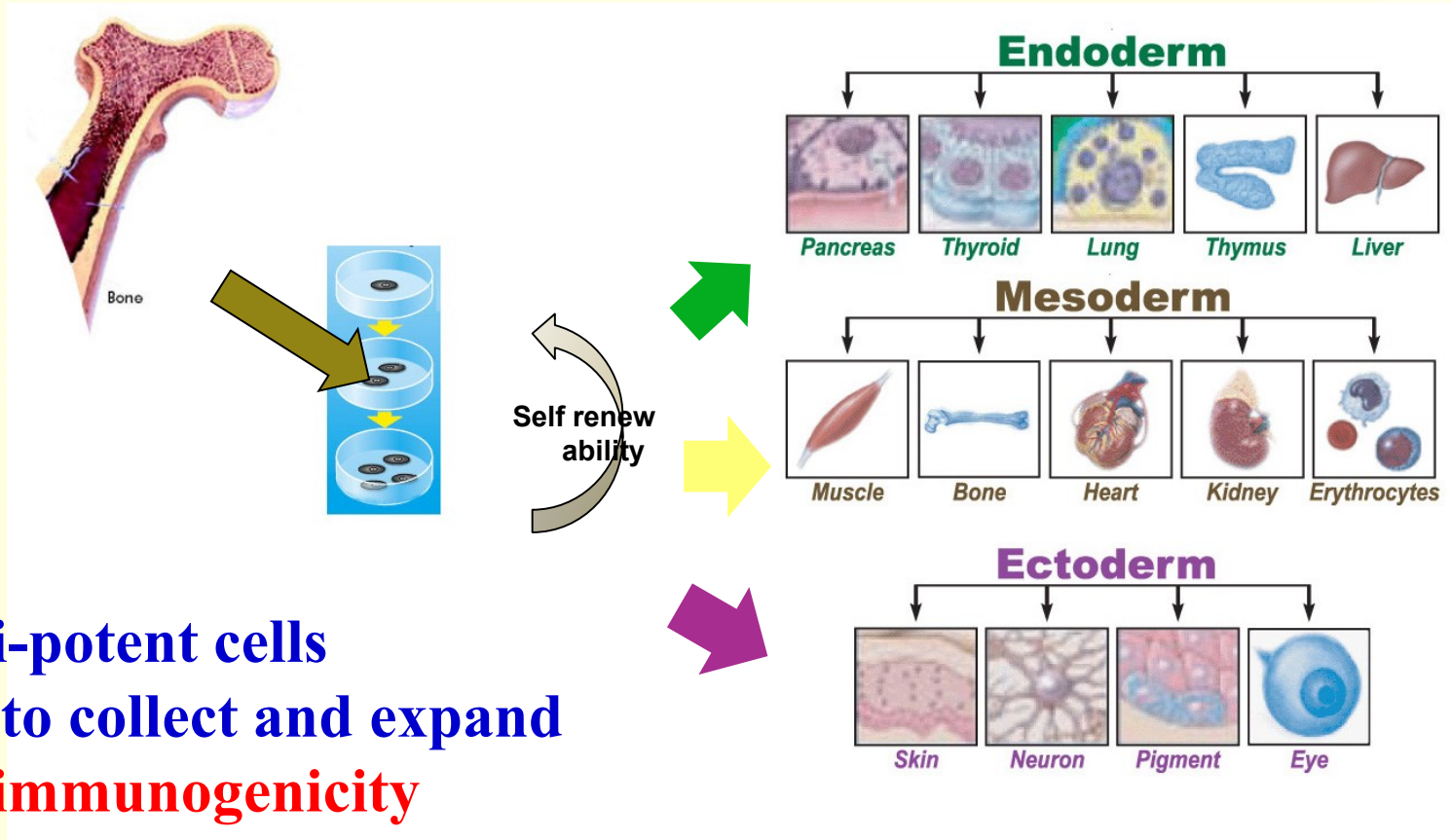
**Dept. of Orthopaedics and Traumatology**

**School of Biomedical Sciences,**

**The Chinese University Hong Kong**



# Bone Marrow Mesenchymal Stem Cells (MSCs)



- Multi-potent cells
- Easy to collect and expand
- Low immunogenicity
- Systemic recruitment
- Home to injury tissues

## **Concise Review: Multipotent Mesenchymal Stromal Cells in Blood**

**QILING HE,<sup>a</sup> CHAO WAN,<sup>b</sup> GANG LI<sup>a</sup>**

<sup>a</sup>Centre for Cancer Research and Cell Biology, Musculoskeletal Education and Research Unit, School of Biomedical Sciences, Queen's University of Belfast, Musgrave Park Hospital, Belfast, United Kingdom; <sup>b</sup>Department of Pathology, Division of Molecular and Cellular Pathology, University of Alabama at Birmingham, Birmingham, Alabama, USA

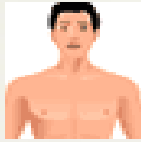
**Key Words.** Peripheral blood • Colony-forming units fibroblastic • Multipotent mesenchymal stromal cells  
Peripheral blood-derived multipotent mesenchymal stromal cells

### **ABSTRACT**

Peripheral blood-derived multipotent mesenchymal stromal cells circulate in low number. They share, most although not all, of the surface markers with bone marrow-derived multipotent mesenchymal stromal cells, possess diverse and complicated gene expression characteristics, and are capable of differentiating along and even beyond mesenchymal lineages. Although their origin and physio-pathological function are still unclear, their presence in the adult peripheral blood might

relate to some interesting but controversial subjects in the field of adult stem cell biology, such as systemic migration of bone marrow-derived multipotent mesenchymal stromal cells and the existence of common hematopoietic-mesenchymal precursors. In this review, current studies/knowledge about peripheral blood-derived multipotent mesenchymal stromal cells is summarized, and the above-mentioned topics are discussed. *STEM CELLS 2007;25:69–77*

## ➤ Progress to date



human



Rabbit



Mouse



Guinea Pig

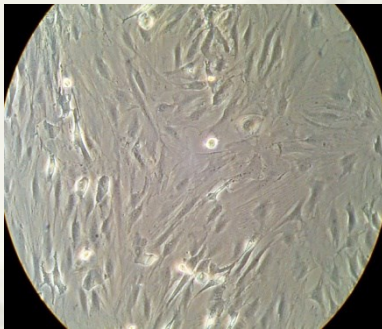
2001. J Cell Biol.

153:1133-1139

**Osteogenesis in vivo; Adipogenesis in vitro**

CD34-/CD45-/CD14-  
CD44+/CD106+ / Type I collagen+  
CD105-/Alkaline phosphatase -/ stromal-

} human cells



**PB-derived  
adherent, clonogenic,  
fibroblast-like cells**

**Circulating osteoblast-lineage cells in humans.**  
New Engl. J. Med. May 12, 2005.

- Sorted osteocalcin+ cells in children
- Formed bone in vivo
- Increased numbers in three adults with recent fractures.



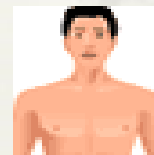
CD34<sup>-/low</sup>  
Osteocalcin<sup>+</sup>

2000. Arthritis Res.

2:477-488

**Osteogenesis in vitro**

CD34-/CD45-/CD14-/CD3-  
CD105+/type I collagen+



human

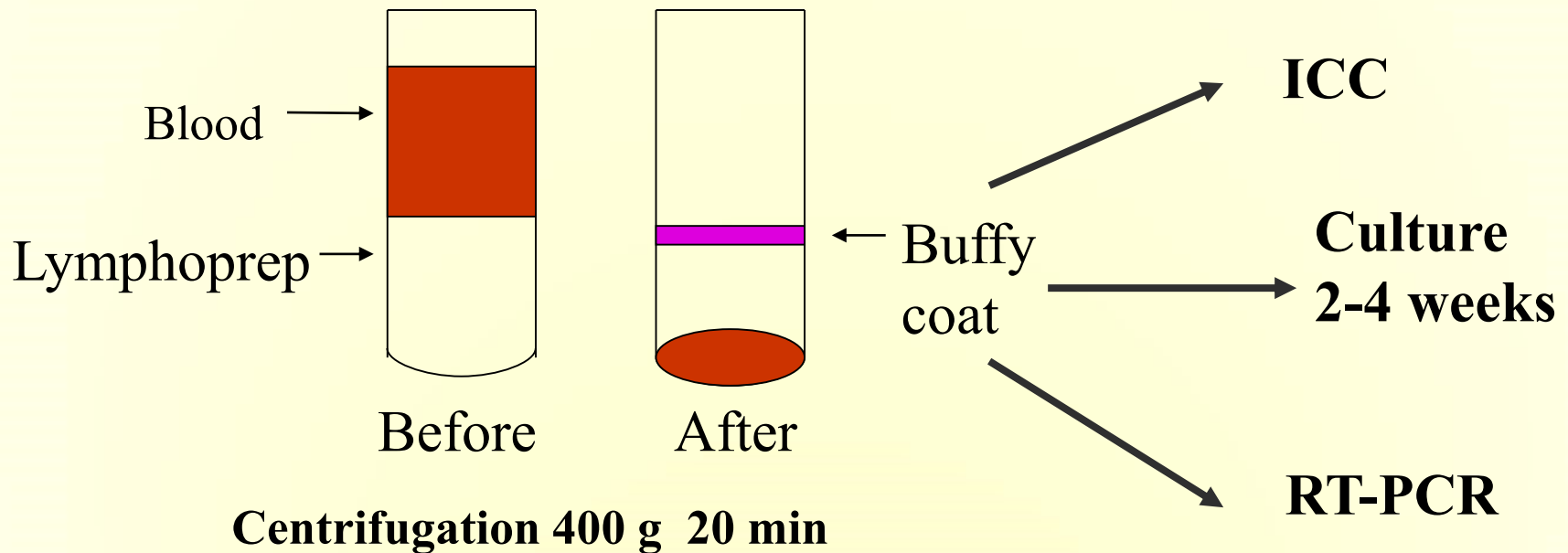
2000. Stem cells

18:252-260

# Study of Circulating MSCs in Fracture Patients

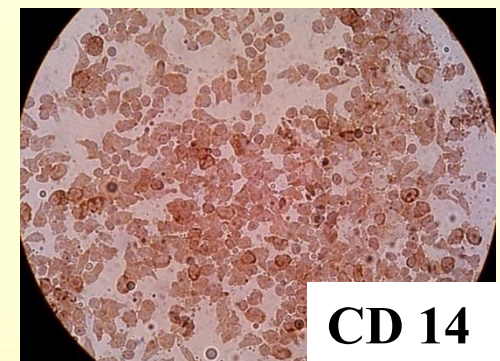
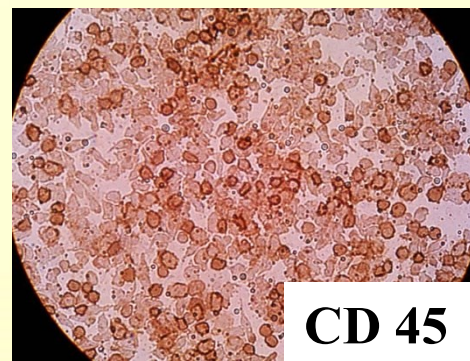
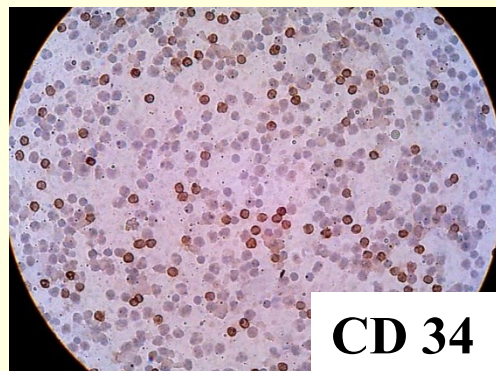
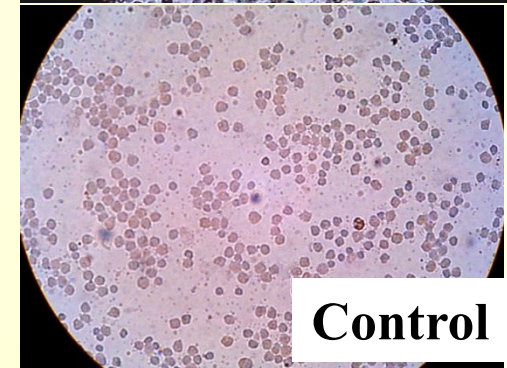
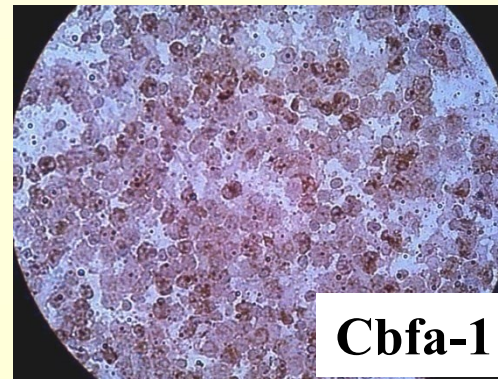
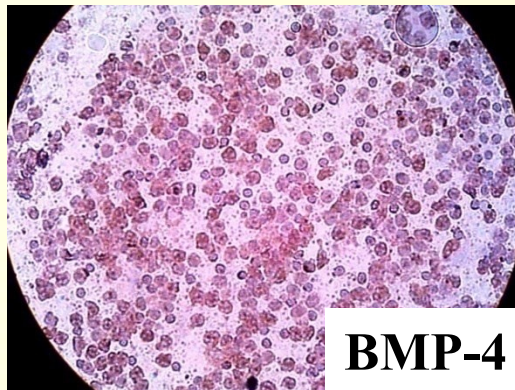
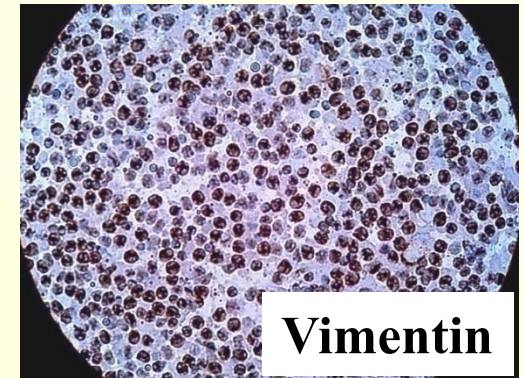
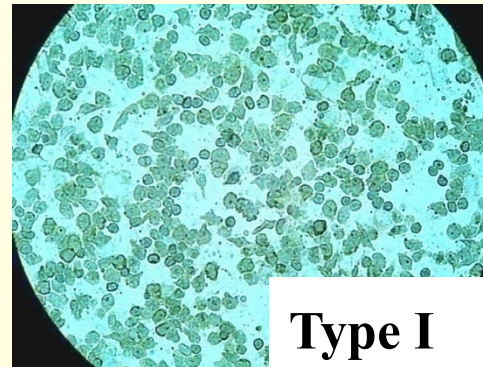
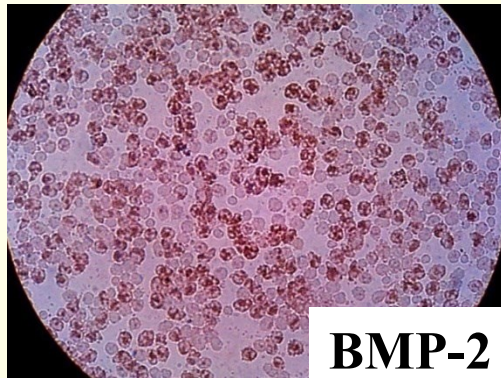
**30 mls of peripheral blood collected from 8 fracture patients, at 3 time-points after fracture ( days 1-3, 9-12 and 16-21) and also from 3 normal volunteers and 3 established non-union**

**Peripheral blood mononuclear cells (PBMNCs) isolated using LymphoPrep™ density-gradient-centrifugation procedure.**



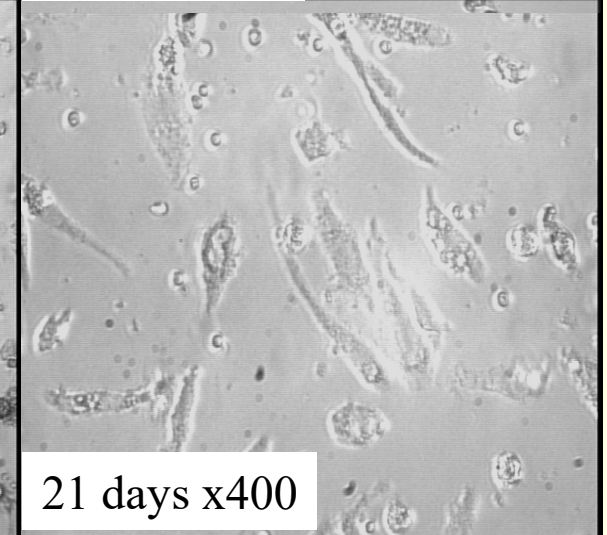
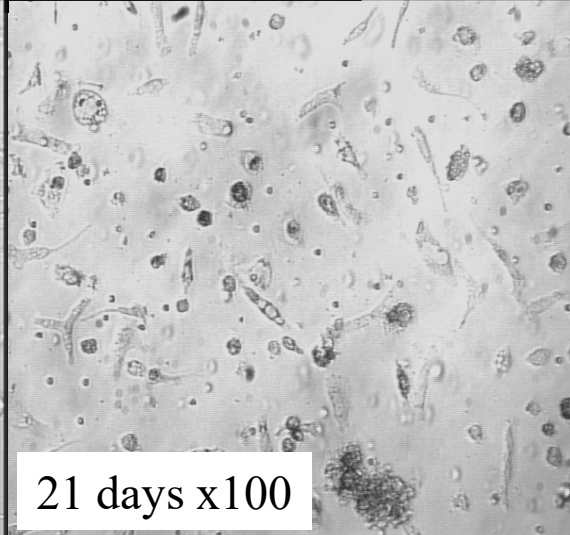
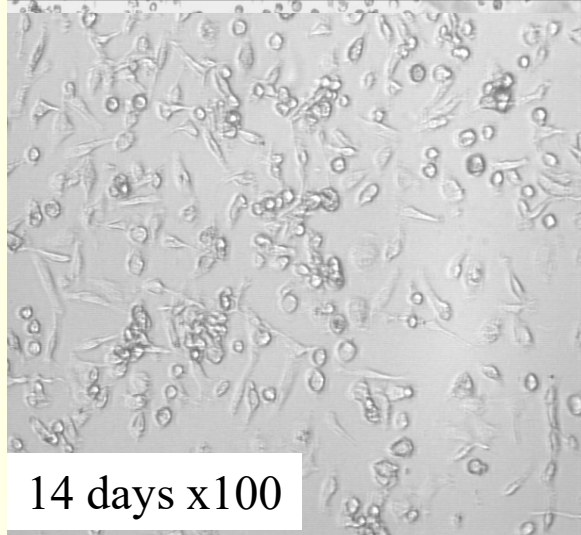
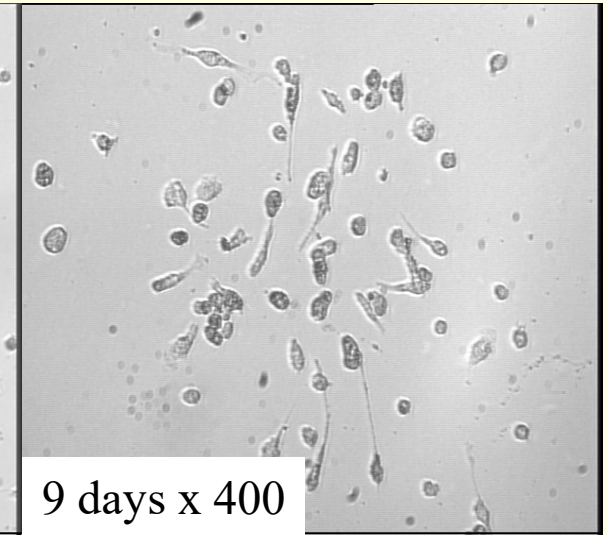
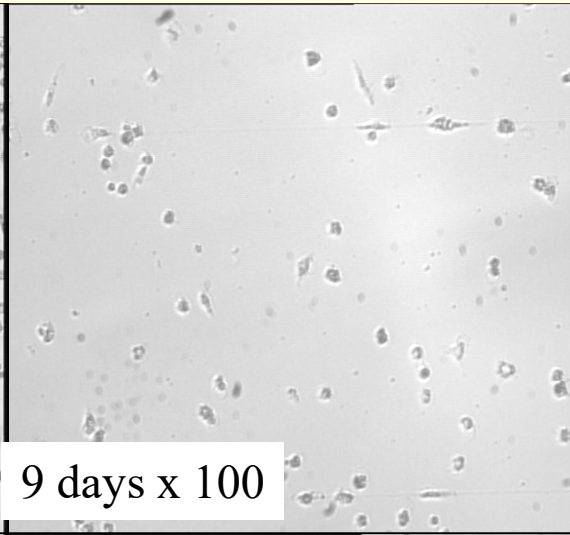
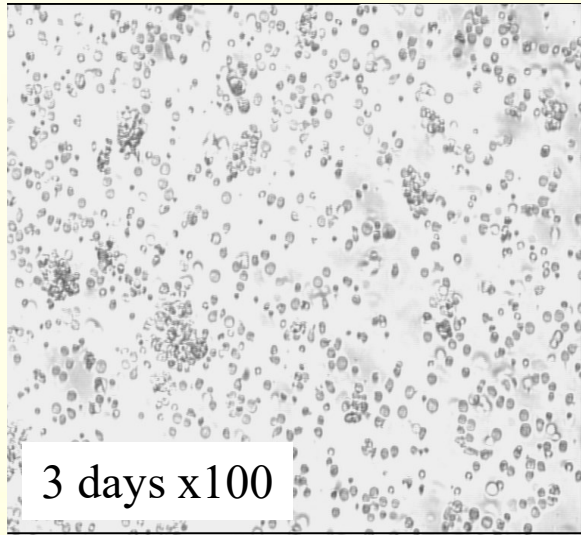


*Immunostaining profile of the PBMNCs from a patient with tibial shaft fracture, at day 3 post-fracture*

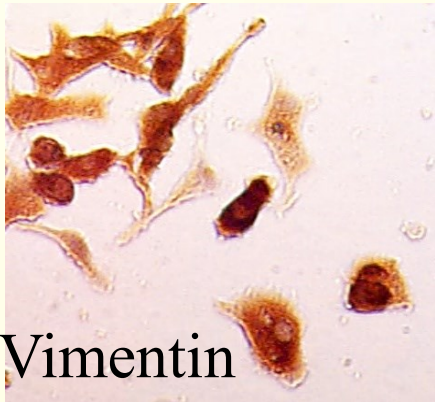




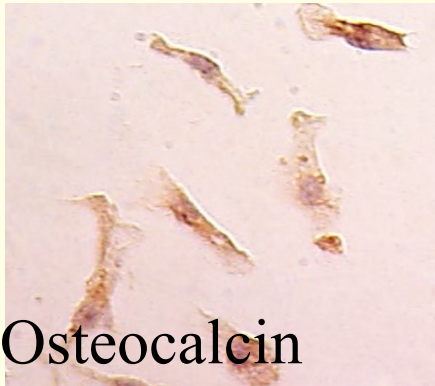
*PBMNCs in Culture with osteogenic medium from a patients with tibial fracture, day 16*



***Immunocytochemistry on a human fracture patient's PBMNC culture at 2 weeks***



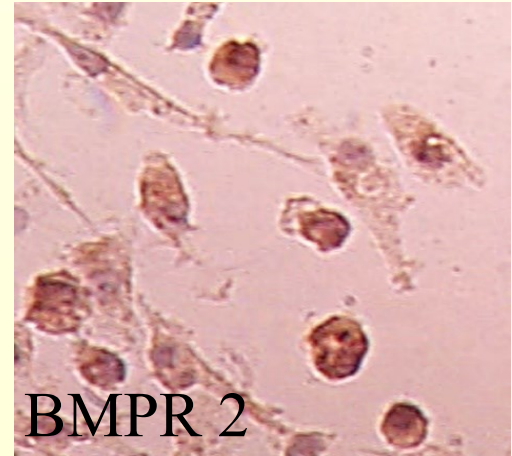
Vimentin



Osteocalcin



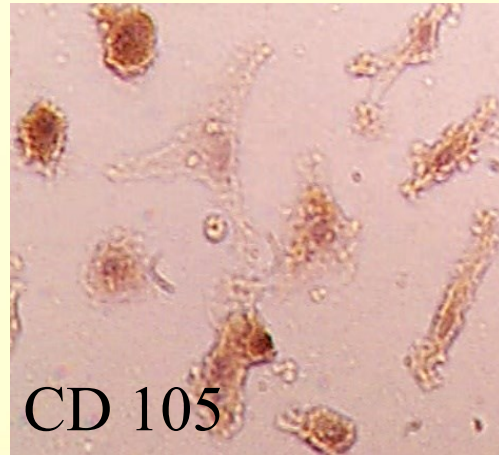
Collagen 1



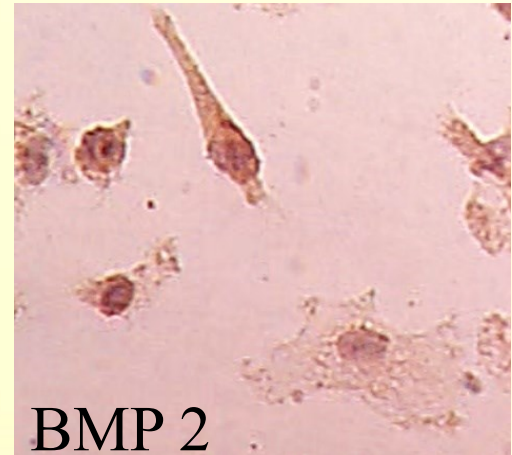
BMPR 2



Negative control



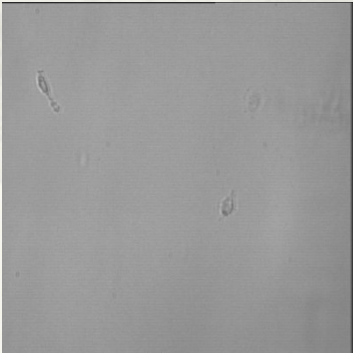

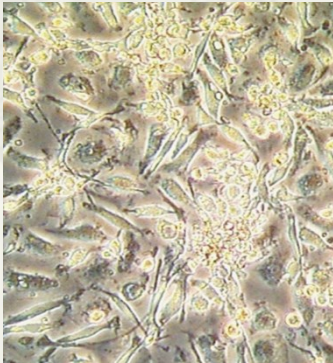
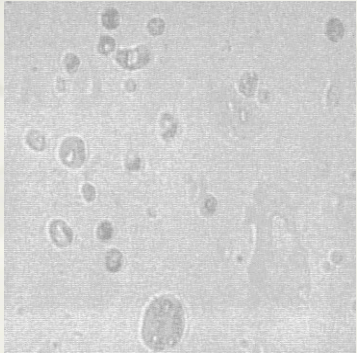
CD 105



BMP 2



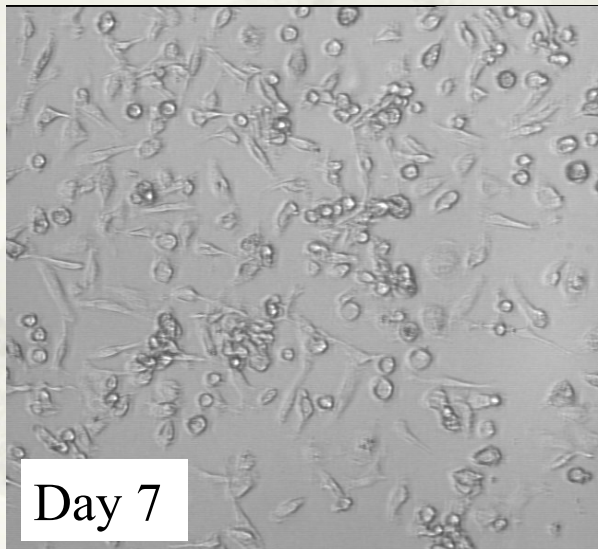
# Summary of cell culture results

	< 4 days Post-fracture	> 14 days Post-fracture	Non-union patients	Control
<b>cases</b>	5	5	3	4
<b>cells</b>	few 	Some 	Many 	None/few 

## ➤ In search of blood borne MSCs

### Normal Adult Peripheral Blood

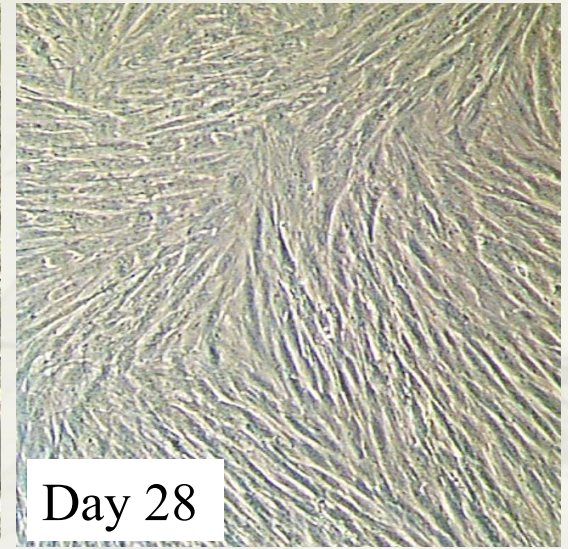
- \* 1 MSC in  $\sim 10^9$  MNCs in normal adult peripheral blood
- \* (vs. 1 MSC in  $10^6$  bone marrow) MNCs
- \* Numbers of MSCs increased in patients with fracture



Day 7



Day 21

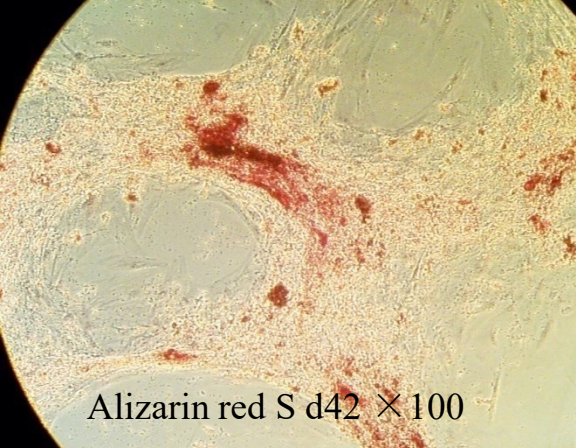


Day 28

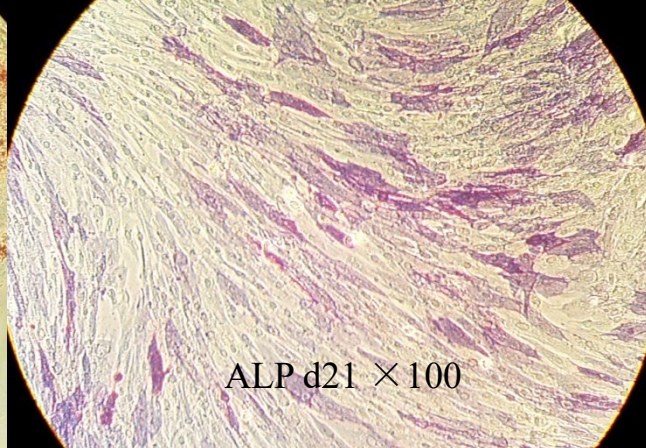
Greater numbers of spindle/polygonal cells found in the peripheral blood MNCs from the patients with fracture non-union, suggesting a systemic recruitment of MSCs may exist (*Shirley, et al. J. Orthop. Res. 2005; 23 (5): 1013-21*)



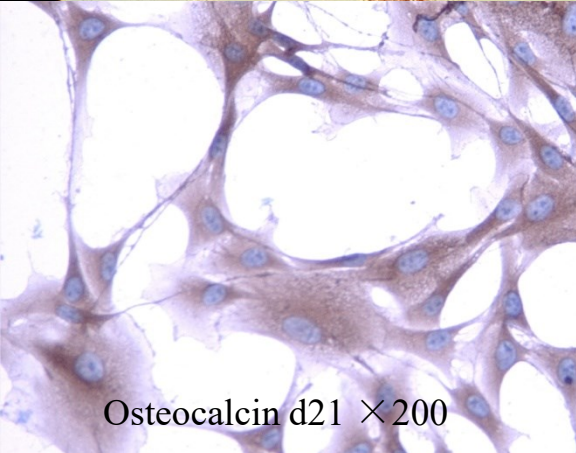
# Characterization of MSCs from non-union patients blood



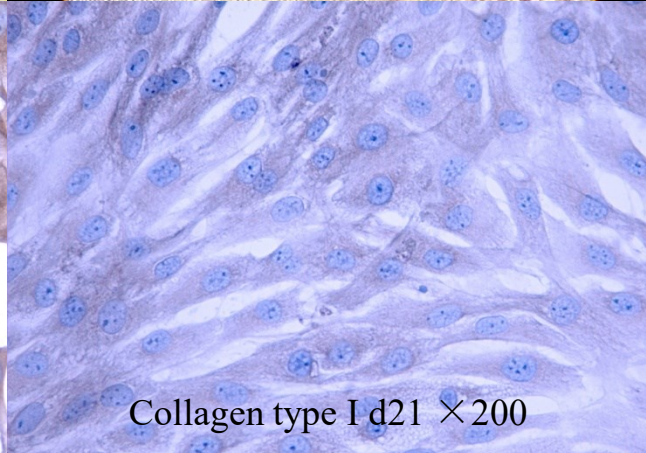
Alizarin red S d42  $\times 100$



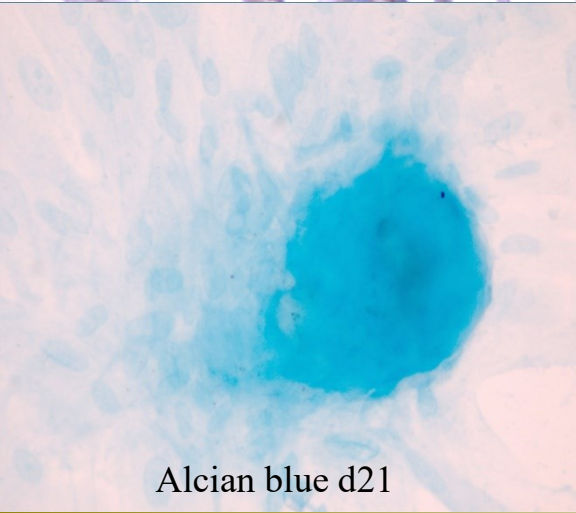
ALP d21  $\times 100$



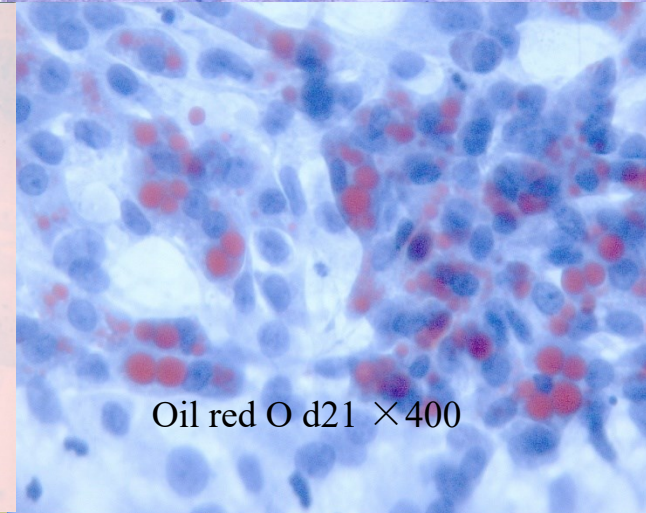
Osteocalcin d21  $\times 200$



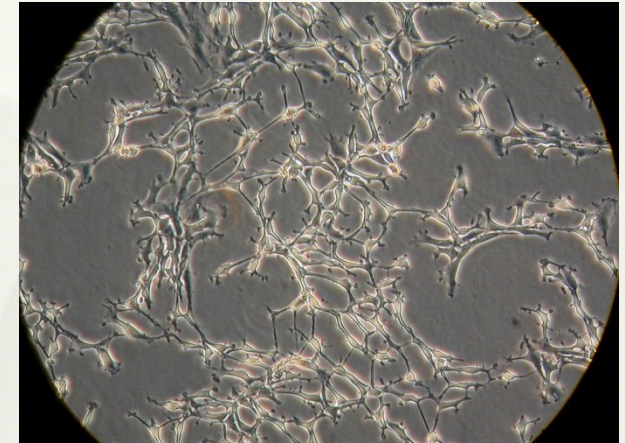
Collagen type I d21  $\times 200$



Alcian blue d21



Oil red O d21  $\times 400$



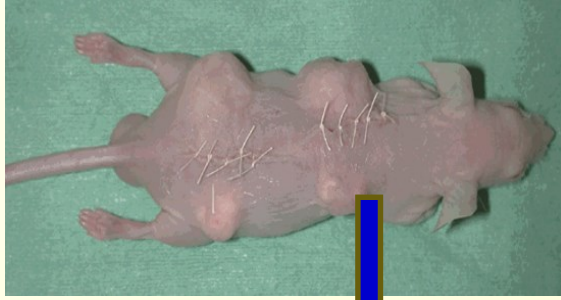
$\beta$ -ME 6h  $\times 100$



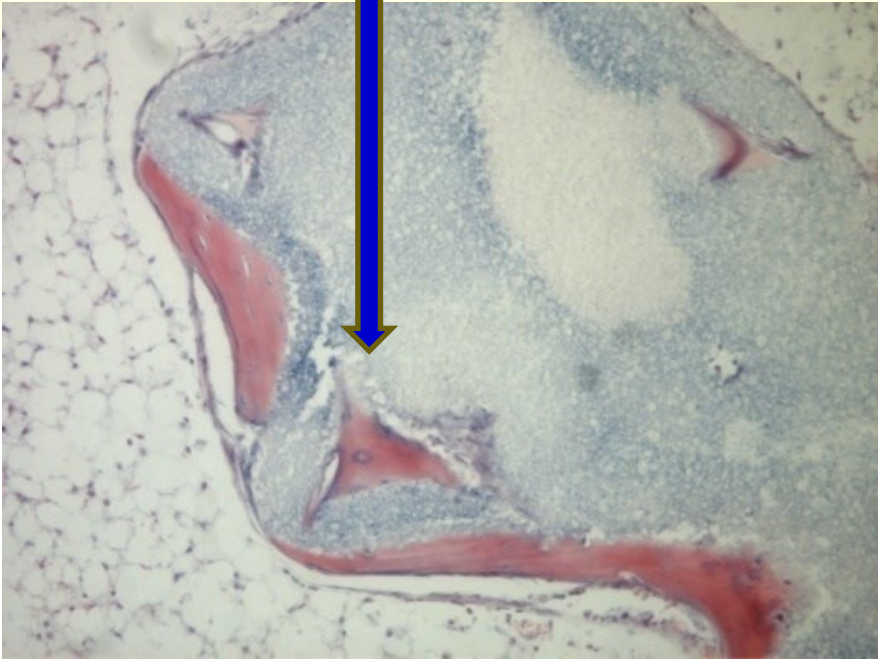
Neurofilament  $\beta$ -ME 6h  $\times 200$



# Differentiation Potential of human blood-borne MSCs

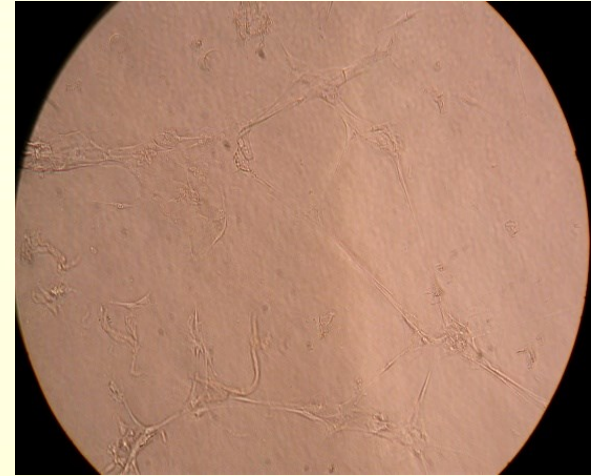


In vivo bone formation study

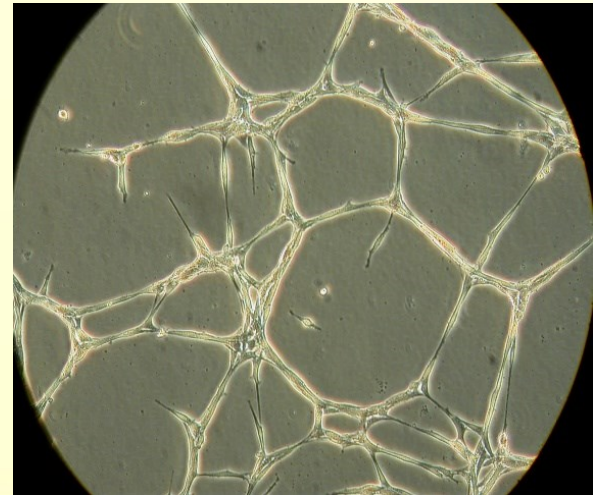


PBMSCsseeded CaP block 3month×50

In vitro angiogenesis



Matrigel 3D culture 24h × 100



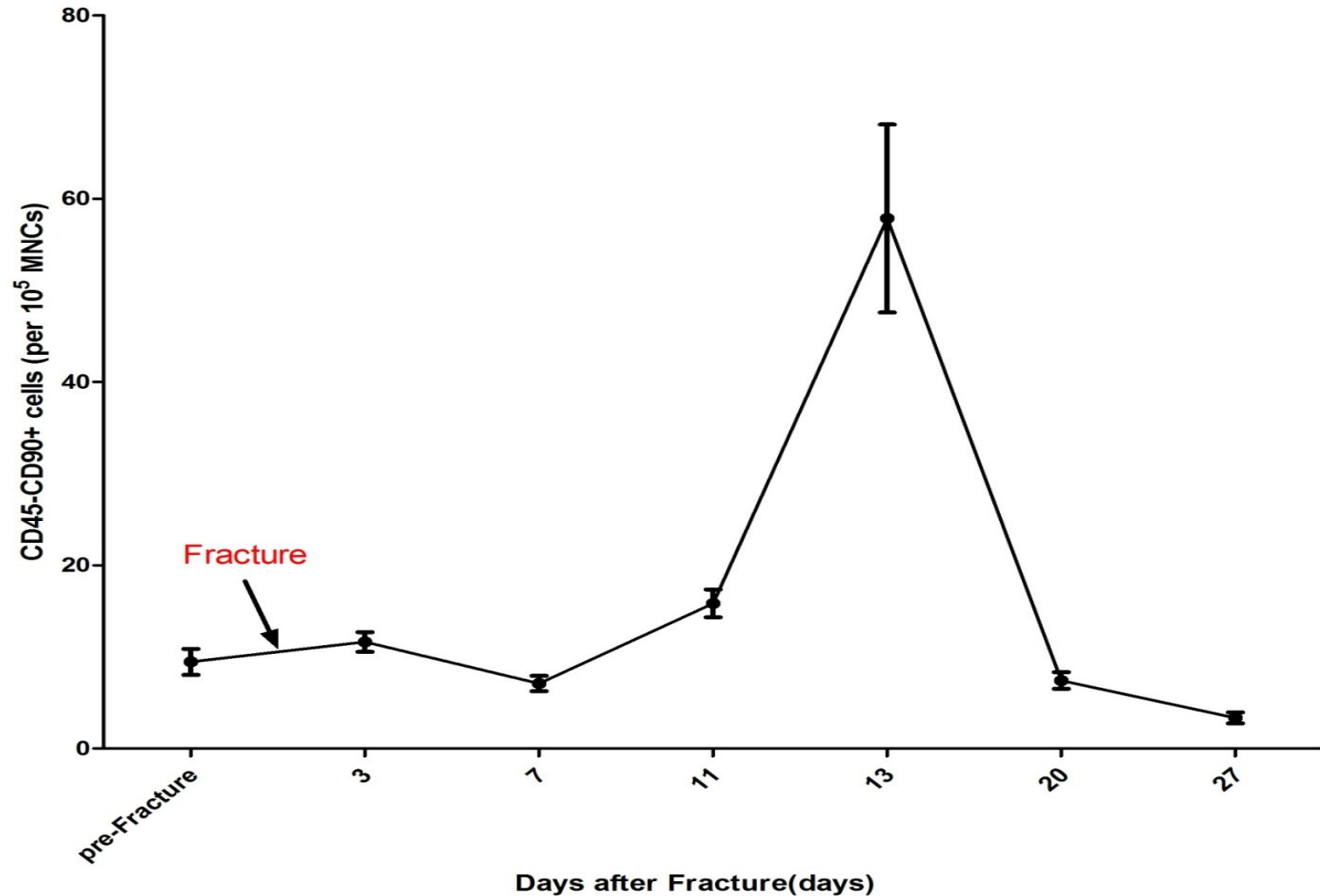
Long term 2D culture 72h × 100

# Change of circulating MSCs during fracture healing in rats

- \* Femoral closed fracture was created in 12 male SD rats (age 12 weeks) with intramedullary nail fixation.
- \* 0.5 ml Peripheral blood was taken from the eye vein at day before fracture, 3, 7, 11, 13, 20, 27 post fracture; CD45 and CD 90 were used to labeled the cells as representative markers for circulating MSCs and subject to flowcytometry analysis.

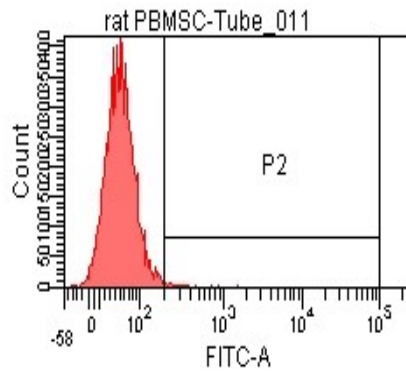


# Results: Changes of blood MSCs (CD45-CD90+) during fracture process

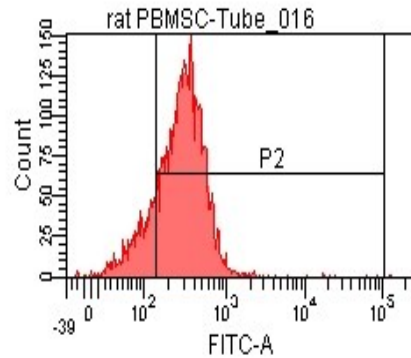




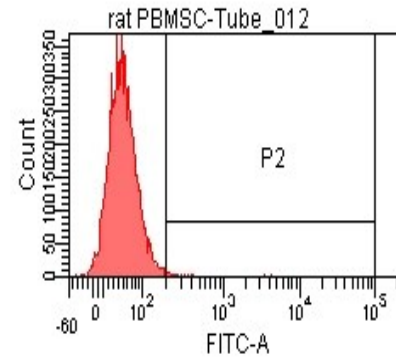
# Results: Characterization and differentiation potentials of Circulating MSCs



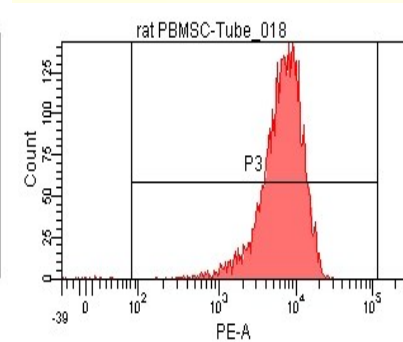
day 8



Day 11



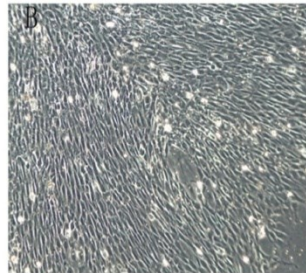
Control



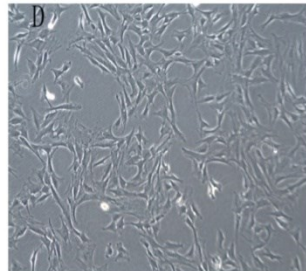
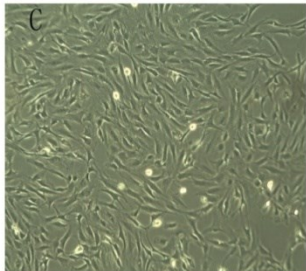
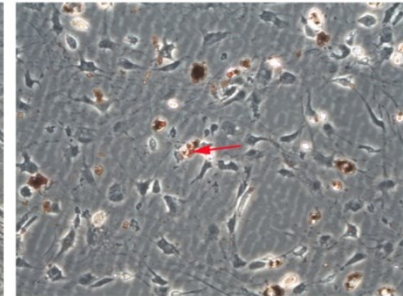
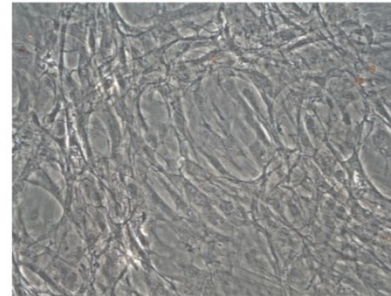
Adipogenesis



P1

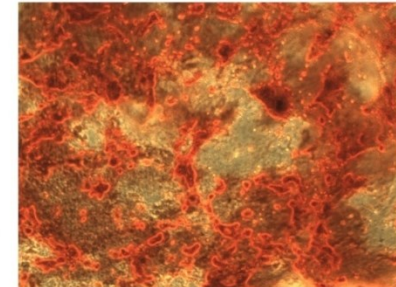
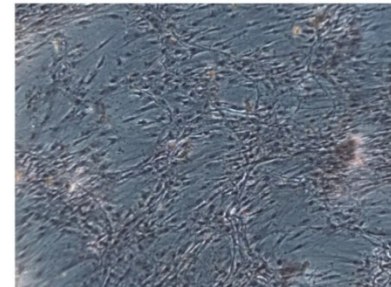


P3

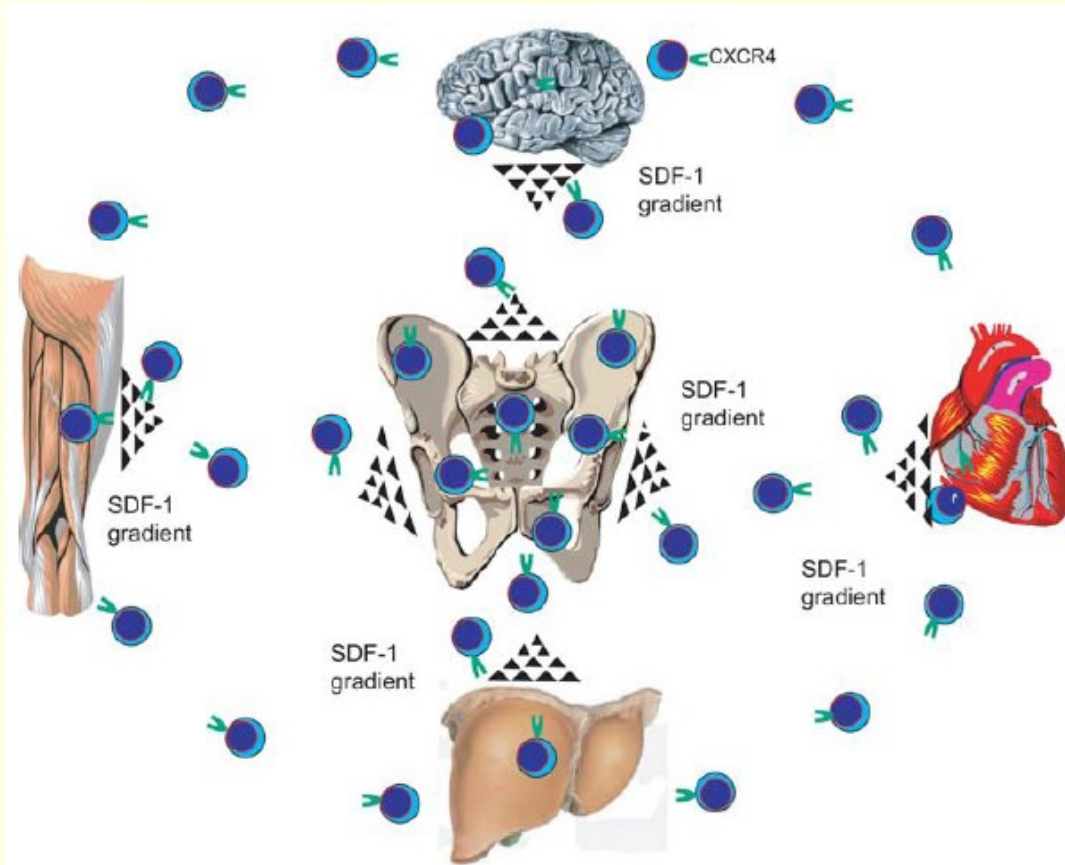


Control

Osteogenesis



# MSCs Home to Injury Sites



MSCs home to a variety of tissues, particularly after tissue injury and ischemia.

Miyahara Y, Nagaya N, Kataoka M, et al. Monolayered mesenchymal stem cells repair scarred myocardium after myocardial infarction. *Nat Med.* 2006 Apr;12(4):459-65.

Carvalho KA, Guarita-Souza LC, Hansen P, et al. Cell Transplantation After The Coculture of Skeletal Myoblasts and Mesenchymal Stem Cells in the Regeneration of the Myocardium Scar: An Experimental Study in Rats. *Transplant Proc.* 2006 Jun;38(5):1596-1602.

Gnecchi M, He H, Noiseux N, et al. Evidence supporting paracrine hypothesis for Akt-modified mesenchymal stem cell-mediated cardiac protection and functional improvement. *FASEB J.* 2006 Apr;20(6):661-9.

Kraitichman DL, Tatsumi M, Gilson WD, et al. Dynamic imaging of allogeneic mesenchymal stem cells trafficking to myocardial infarction. *Circulation.* 2005 Sep 6;112(10):1451-61.

# *Where do circulating MSCs come from?*



Journal of Orthopaedic Research 23 (2005) 1013–1021

---

Journal of  
Orthopaedic  
Research

---

[www.elsevier.com/locate/orthres](http://www.elsevier.com/locate/orthres)

## Systemic recruitment of osteoblastic cells in fracture healing

Denise Shirley <sup>a</sup>, David Marsh <sup>a</sup>, Grant Jordan <sup>a</sup>, Stephen McQuaid <sup>b</sup>, Gang Li <sup>a,\*</sup>

<sup>a</sup> *Department of Trauma and Orthopaedic Surgery, School of Medicine, Queen's University Belfast,  
Musgrave Park Hospital, Belfast BT9 7JB, UK*

<sup>b</sup> *Department of Pathology, Royal Victoria Hospital, Belfast BT12 6BJ, UK*

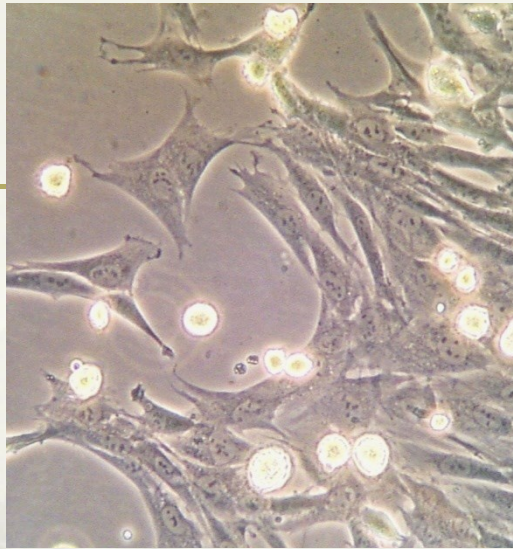
Accepted 28 January 2005



# MSCs homes to fracture sites through peripheral circulation



Bone marrow harvested



Rabbit bone marrow  
MSCs culture

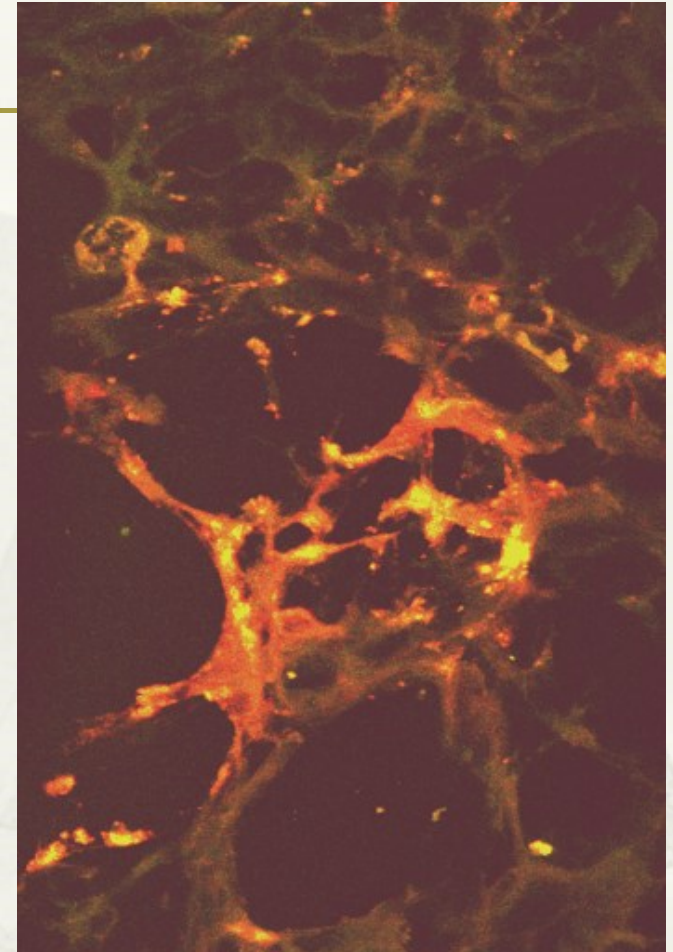
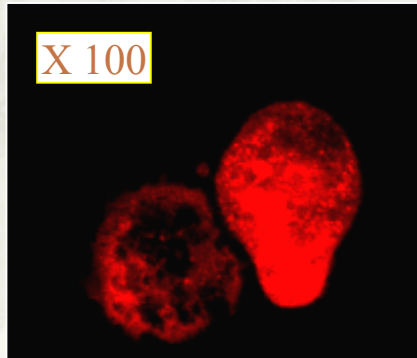
## Cell Labeling

PKH26 Red Fluorescent Cell Linker Kit  
For general cell membrane labeling  
Product Code: PKH26-GL

**SIGMA**

Cell Biology

X 100



*Shirley D, et al, Journal of Orthopaedic Research, 2005, 23 (5): 1013-21.*

# Re-implantation

In each group some animals were sacrificed at 3 & 12 weeks



48 hours  
post Fx

Culture  
3 weeks

Control E

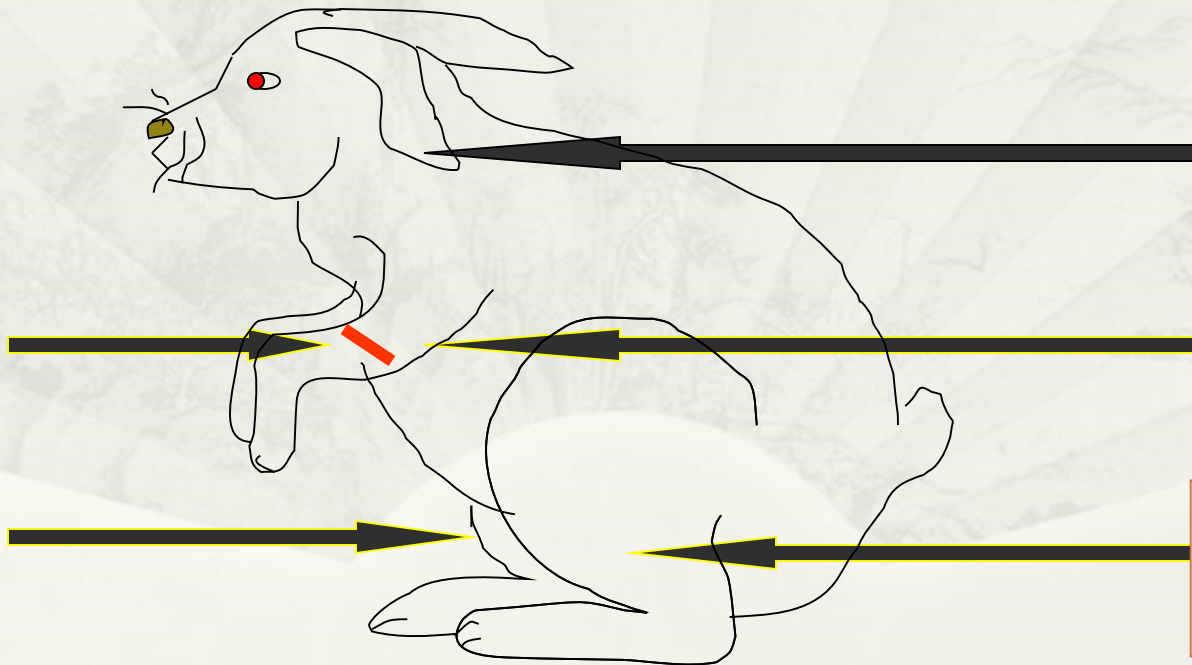
Ulnar  
defect

Ear vein B

BM  
harvest

Fracture  
site A

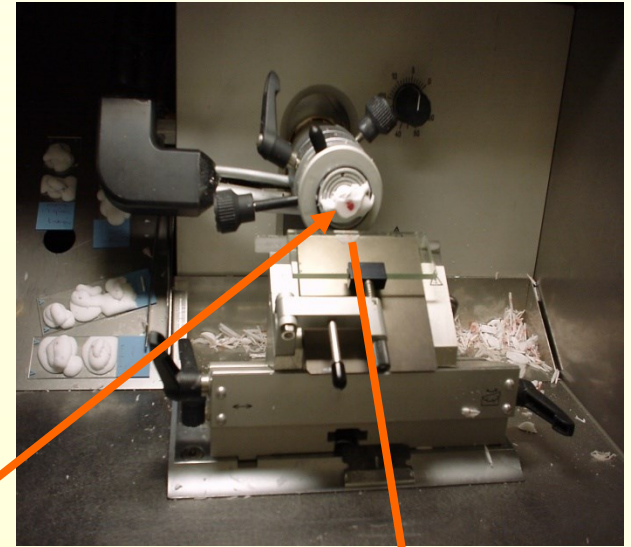
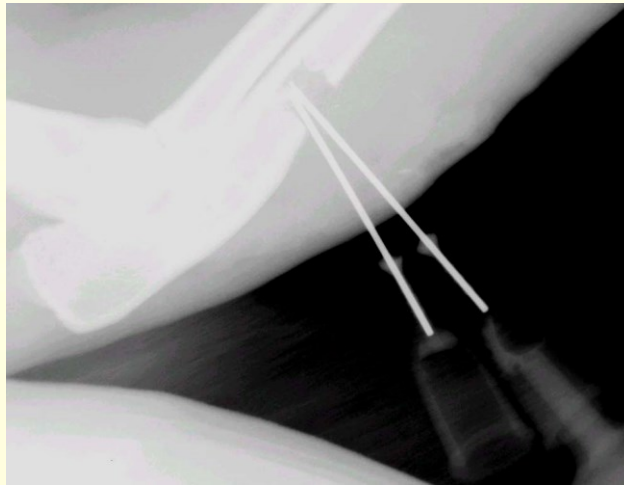
Remote  
BM site C



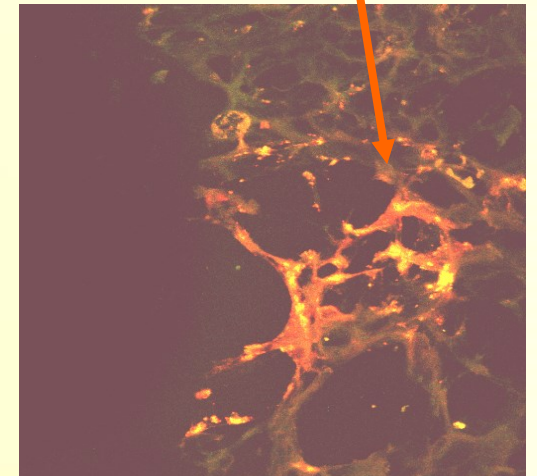
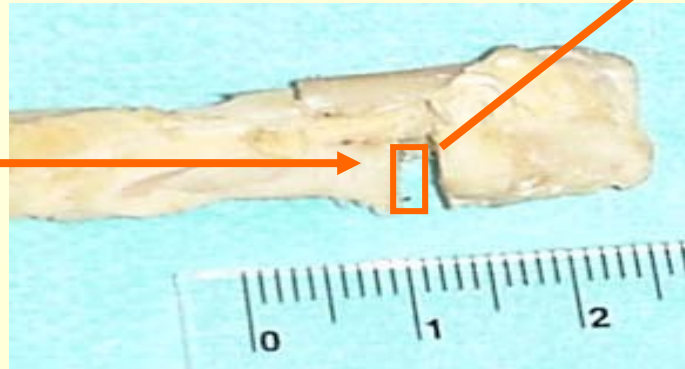
*Shirley D, et al, Journal of Orthopaedic Research, 2005, 23 (5): 1013-21.*

# The tissues retrieved for frozen section – (5 $\mu$ m)

Animals were sacrificed at 3 and 12 weeks after cell implantation



Gap tissue

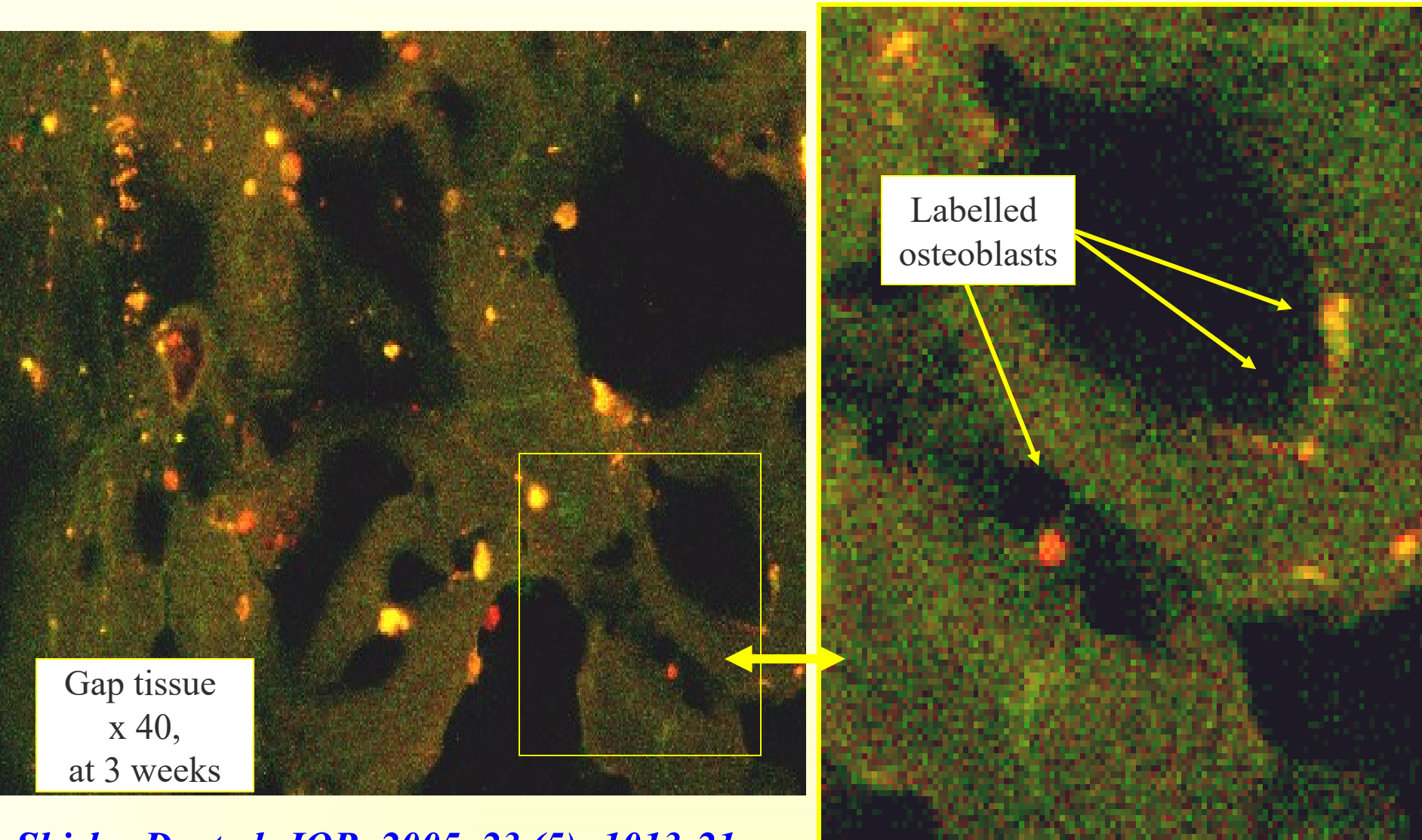


- Liver, lung, kidney, and spleen,
- Also cytopins of BM and blood

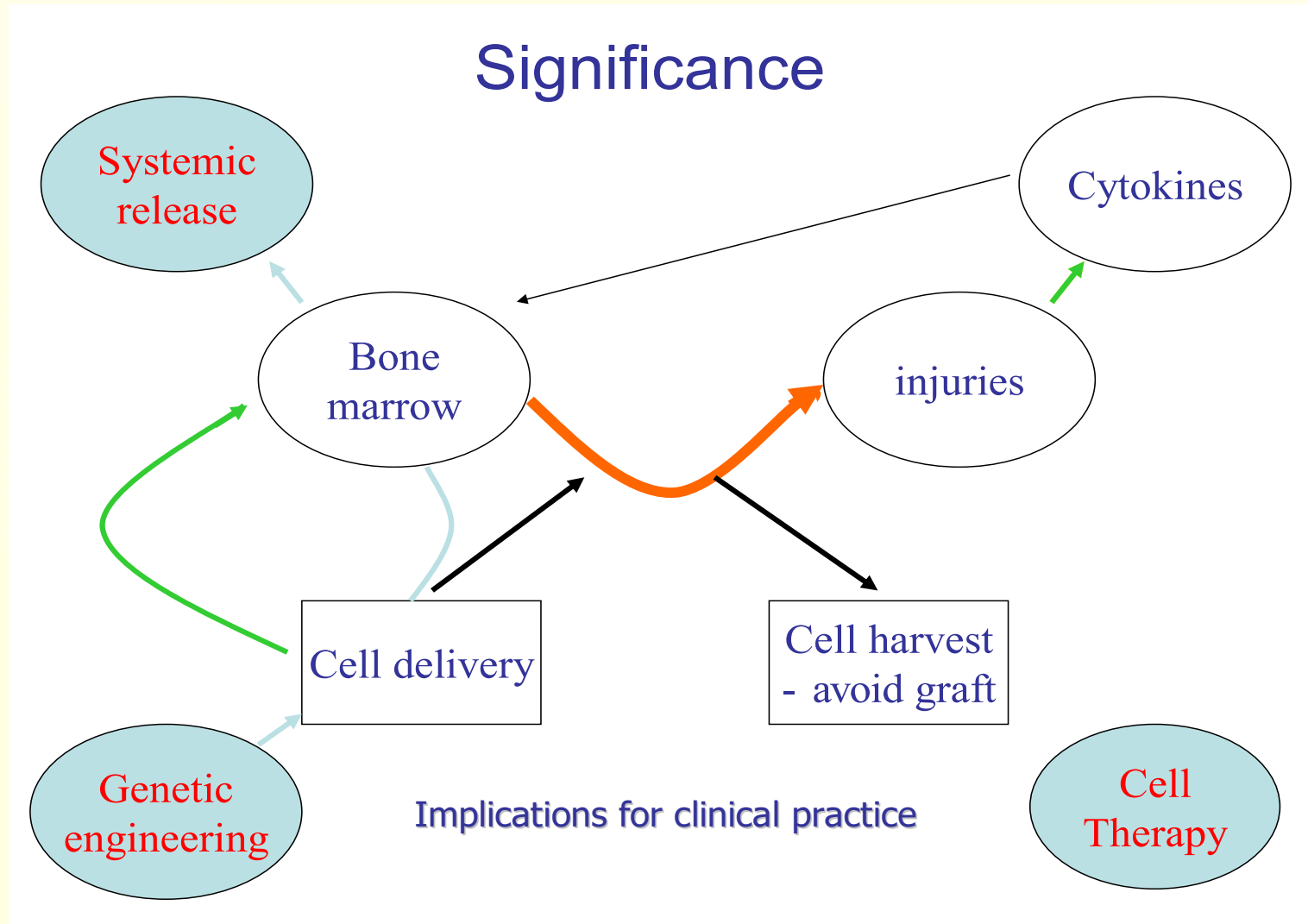
(representative samples only)



# Labelled cells from remote marrow identified at the fracture gap ( Group with systemic injection of allogenic MSCs )



- **Some osteoblasts integral in fracture repair come from remote bone marrow sites.**
- **They were actively recruited through the peripheral circulation.**



# **Local Vs. Systemic MSCs Administration**

- **Local injection of autologous MSCs have been shown to promote fracture healing (Chanda et al, 2010; Li, et al 2010) .**
- **3-4 weeks time is needed to culture-expend MSCs to sufficient therapeutic numbers, may miss the “window of opportunities”.**

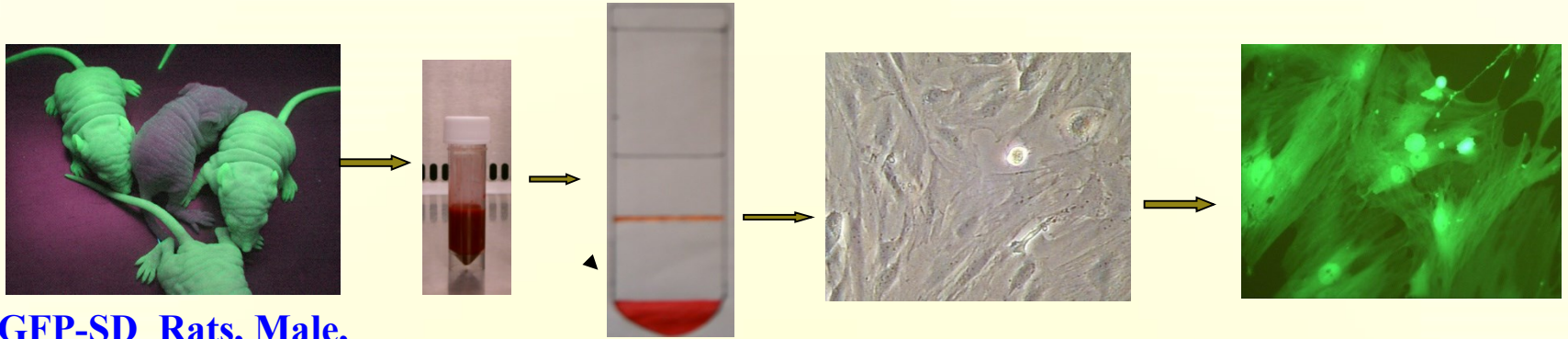


## **Local Vs. Systemic MSCs Administration**

- **Locally delivered MSCs often face hostile microenvironment: lack of blood supply; infection; inflammation that minimize their survival and impair their function *in vivo*.**
- **Systemic administrated MSCs may reach the fracture sites through circulation, where the sufficient blood supply will enhance their survival and function.**

# Materials and Methods

## Cell Preparation



GFP-SD Rats, Male,  
3-6 month old

- ❖ Isolation of BM-MSCs and skin fibroblasts from GFP-Rat
- ❖ Flow cytometry analysis for cell surface antigen markers:
  - Positive: CD44, CD73, CD90, CD146
  - Negative: CD31, CD34, CD45
- ❖ Differentiation assays: adipogenesis, osteogenesis, chondrogenesis

# Materials and Methods

## Animal Experimental Groups



- ❖ 48 male SD rats (age: 12 weeks) had right femoral closed fracture
- ❖ Fracture was fixed with intramedullary nail
- ❖ Animals were randomly assigned into 4 experimental groups (n=12)

### **PBS Heart Injection Group (Control)**

0.5ml PBS/ Rat was given at 4 days post-fx

### **MSCs Heart Injection Group**

$2 \times 10^6$  GFP-MSCs in 0.5ml PBS/ Rat was given at 4 days post-fracture

### **Fibroblast Heart Injection Group**

$2 \times 10^6$  GFP-Fibroblasts in 0.5ml PBS/ Rat was given at 4 days post-fracture

### **MSCs Fracture Site Injection Group**

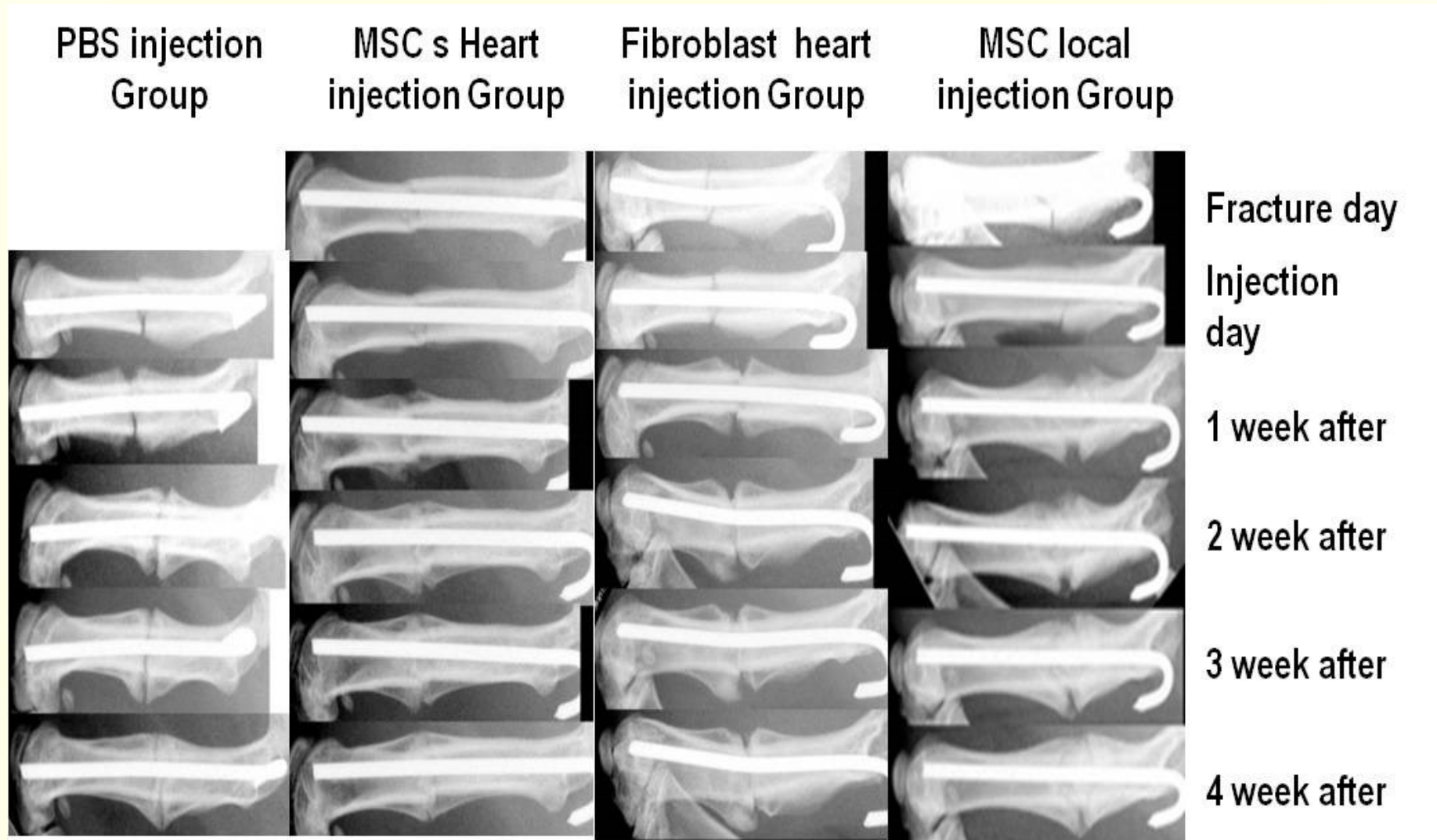
$2 \times 10^6$  GFP-MSCs in 0.5ml PBS/ Rat was given at 4 days post-fracture



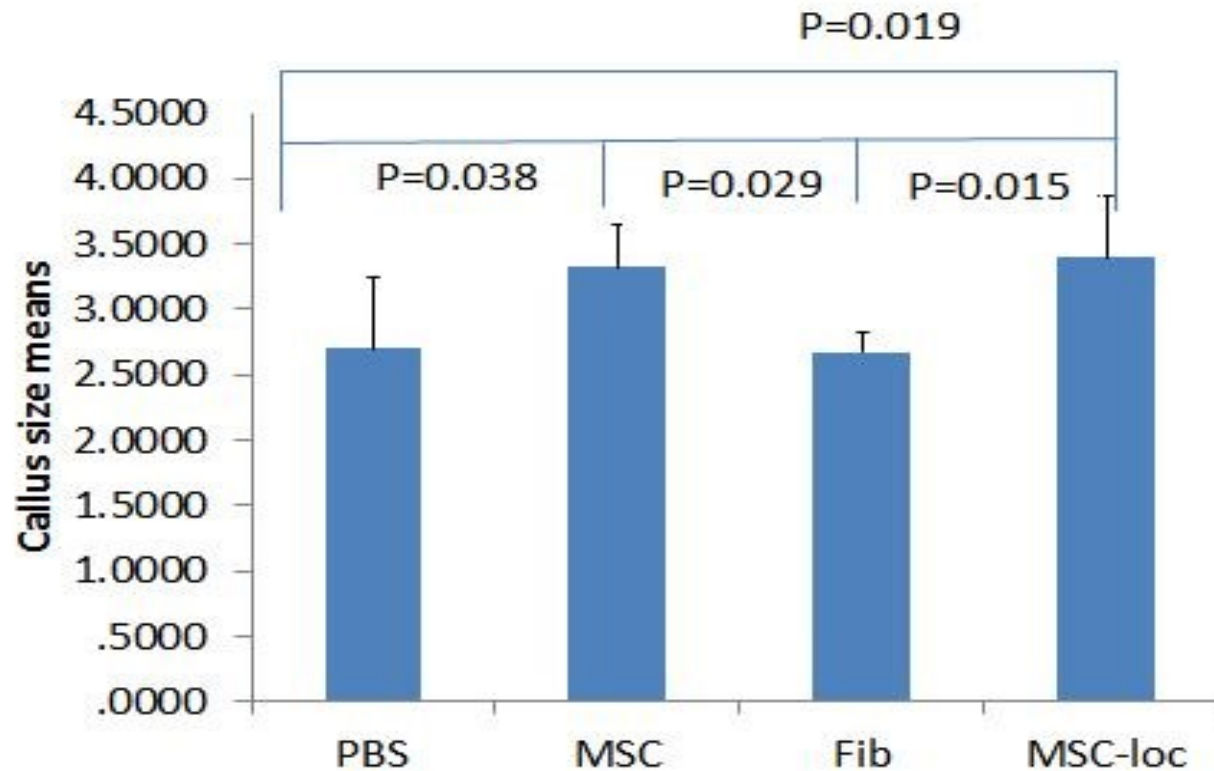
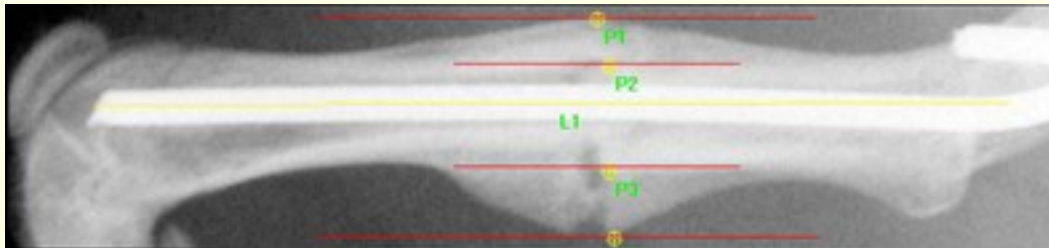
# Outcome Measurements

- **Weekly body weight and X-ray .**
- **Terminated at 5 weeks post fracture, both femurs were harvested.**
- **Micro-CT examination followed by four-point bending mechanical testing.**
- **Histology and immunohistochemistry examinations.**

# RESULTS: X-Ray



# RESULTS- COMPARING THE SIZE OF THE CALLUS

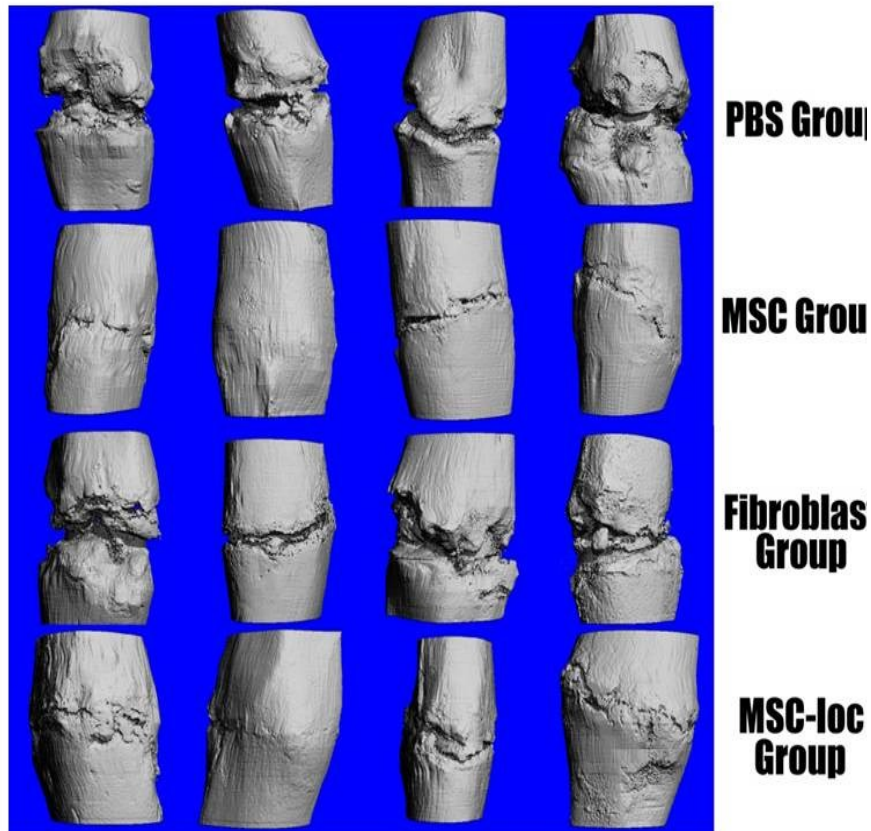




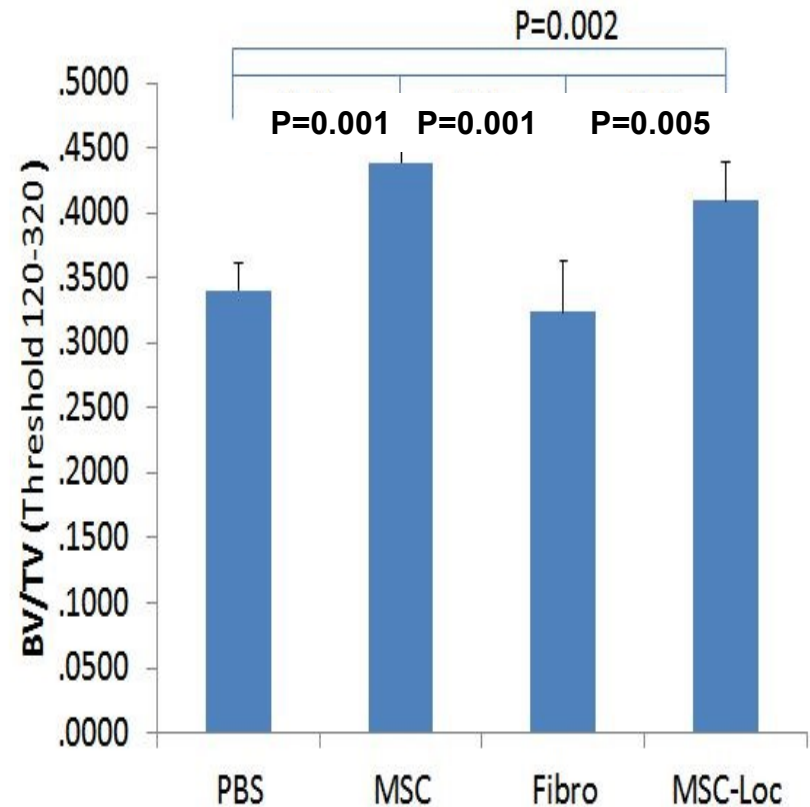
# RESULTS: MICRO CT ANALYSIS

## 3D Reconstruction

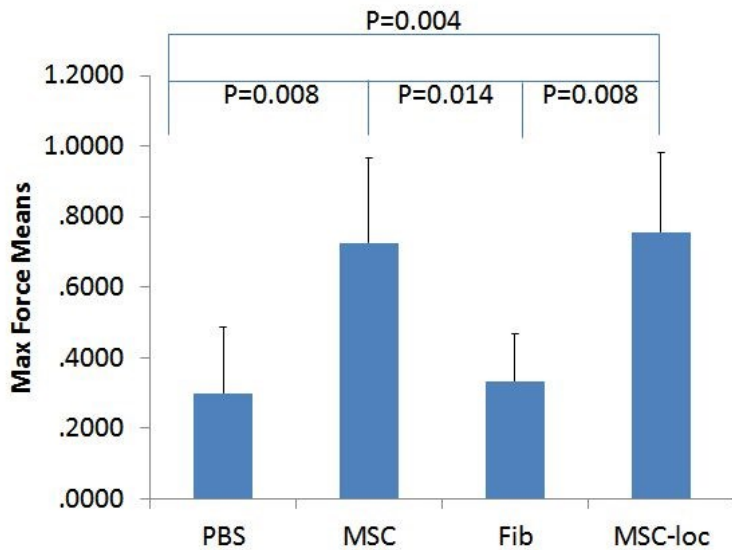
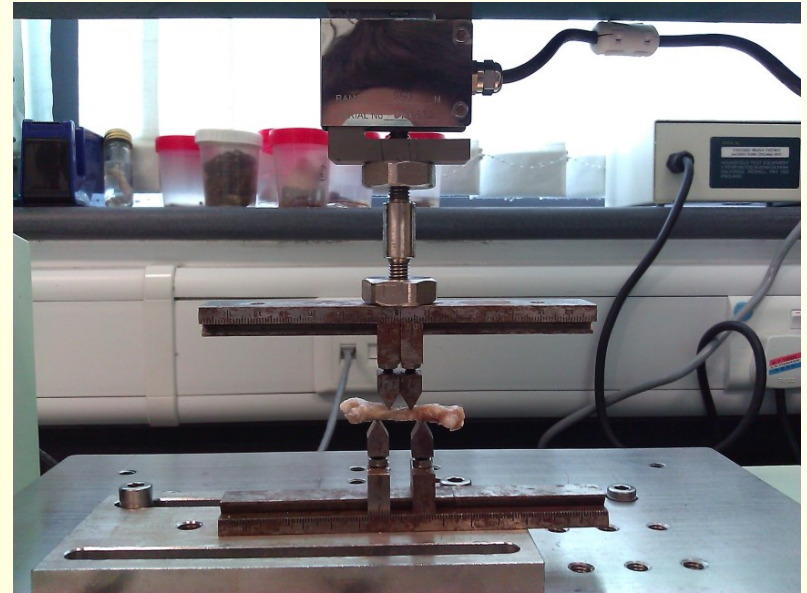
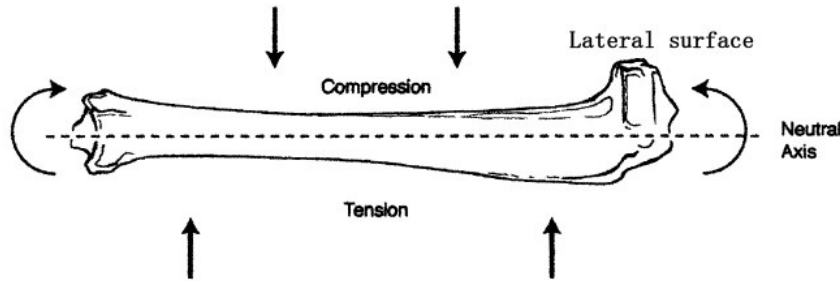
Animal 1   Animal 2   Animal 3   Animal 4



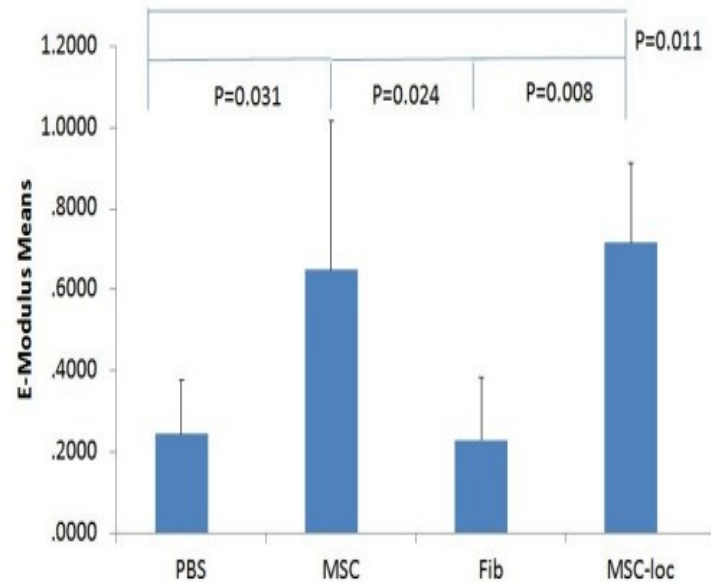
## Bone Volume / Total Volume (BV/TV)



# Results: Four-point Bending Mechanical Testing



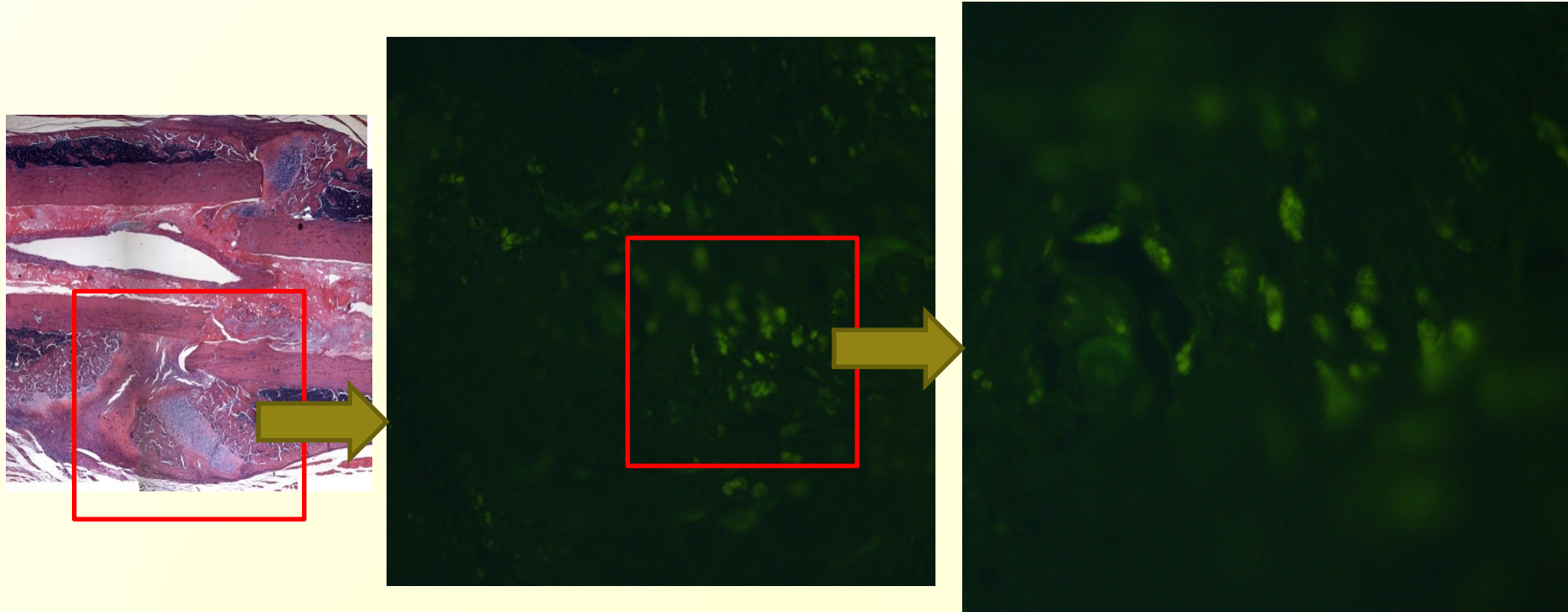
Max Force



E-Modulus (Stiffness)

## Results: Histology & Immunofluorescence for GFP-MSCs

### MSC systemic injection group



**GFP-positive cells were found at the fracture gap 4 weeks following the systemic GFP-MSCs injection.**



# SUMMARY

- Both systemic and local injection of allogeneic MSCs promoted bone fracture healing, through enhancing the callus size and biomechanical properties.
- The MSCs were present at the fracture site and participated in fracture healing at 4 weeks following their systemic injection. The underlying mechanisms need further investigations.

# CONCLUSION

- Our findings provide insight for developing systemic administration of allogenic MSCs as a novel therapy strategy for patients with poor fracture healing conditions, such as multiple or high-energy fractures.



**Acknowledgement of Funding Supports:**  
**HK RGC, HK ITF, NSFC, Shanghai Gov.,  
Shenzhen Gov., Amgen USA, Eli Lilly USA,  
China 973 Project, SMART-IIM**

